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Indolocarbazoles. 4. Synthetic Studies Towards Staurosporine and Tjipanazoles: Reactions of Indolocarbazole with Glycals.

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Abstract: In an attempt to construct the unique N,N'-bidentate glycosyl linkage found in the Staurosporine 1 class of natural products, the first example of an acid catalyzed 2,6-condensation of an activated pyran with indolocarbazole 4 is reported. In pursuit
of the real system, a variety of glycals were reacted with indolocarbazole 4. Formation 1.3-connections along with the expected product (A) and its' synthetic transformation to a potentially useful intermediate 24 are detailed.

The Indolocarbazole natural products possess interesting biological activity and are the subject of a recent review article.1 Noteworthy examples include the kinase inhibitors Staurosporine **1** and K252a 2, and the antifungal Tjipanazoles 3. As a part of our ongoing involvement² with synthesizing analogs of these natural products as anti-tumor agents³ we wished to develop a general synthesis of pyranosylated indolocarbazoles especially with the two glycosyl bonds typical of **1.** After having failed to effect ring expansion on a synthetic K252a analog₄ we were attracted to the potential of glycals as an entry into pyranosylated indolocarbazoles.

Several routes to the aglycones are known,⁵ but the synthesis of the complete skeleton of staurosporine was only achieved very recently by Danishefsky et. al.⁶ For our own studies, we chose to exploit the increased nucleophiicity of the parent heterocycle 4, since we had established an efficient protocol for subsequently installing the crucial imide/lactam system $(7 -\epsilon)$ **la**).² Herein, we report the reactions of 4 with unsaturated sugars.

Reterosynthetieally, the two related routes shown in Scheme 1 seemed attractive.

Our initial exploration of the "intramolecular" route is shown in Scheme 2. The first glycosidic bond formed efficiently, providing 8 as an anomeric mixture. The clear equatorial preference for the heterocycle. evident from PMR, was reflected in the reluctance of the final closure. Under the indicated conditions, we were able to obtain a modest yield of 9, the parent system of staurosporine.

Scheme 2: Synthesis of staurosporine parent system.

Recognizing that the glycal derived intermediates might be more stable, we proceeded to carry out our proposed intermolecular bis glycosidation. Bis enol ether 5 was synthesized in an efficient manner from tri-Oacetyl-D-glucal using a combination of literature procedures.^{7,8} Remarkably bis enol diacetate 5 was very stable and was resistant to a number of glycosidation attempts with indolocarbazole under different lewis acid conditions.

We next turned our attention to the intramolecular strategy. The plan was to glycosidate indolocarbazole with tri-0-acetyl-D-glucal 10 and then unveil the second enol bond from the acetoxy methyl group of 10A for electrophilic exo cyclization. Glycals normally react with nucleophiles to give either 2-deoxy glycosides by addition to the vinyl ether bond, or 2,3-dideoxy-hex-2-enopyranosides via the Ferrier rearrangement.9 However, when indolocarbazole reacted with tri-O-acetyl-D-glucal in the presence of protic acid (PTSA) in methylene chloride at room temperature, the expected product was formed only in trace amounts. The major products were cyclic diasteroisomers 10B and 1OC. Such anomalous C-3 substitutions on glycal systems are observed¹⁰ only under high temperature conditions which promote sigmatropic or allylic rearrangement and such factors could not be invoked here. This prompted us to further elaborate this observation by treating indolocarbazole with other glycals¹¹ and the results are summarized in Scheme 3. The overall yields are unoptimised and are modest due to the capricious dimerization of these glycals¹² under the reaction conditions. Improvement was observed when the glycal was added by syringe pump over an extended period as a solution. The product distribution between the simple desired glycosidated product (A) and the unusual diasteroisomeric products $(B + C)^{13}$ is reversed in the case of arabinal and galactal acetates (entry 13 and 14). A tentative explanation of our results could be a possible involvement of an allylic carbonium intermediate 16¹⁴ whose formation is inhibited by the cis orientation of the acetates at C-3 and C-4.

Similar observations were made when indolocarbazole reacted with 2,5-dimethoxy dihydrofuran 17. The main product obtained was 19 via an analogus allylic carbonium intermediate 18. The

assignment of the products was derived unambiguously from the characteristic coupling Constants and verified by the upfield shift of the acetate methyls syn to the indolocarbazole ring, due to anisotropy.¹⁵

These observations restored **OUT** original plan of transforming an intermediate like **14A to the exo**cyclic enol 24. This w a s accomplished in

steps via 23. 23 can also be obtained alternatively from readily accessible 20.7 Intermediate 21 can be viewed as an analog of *Tjipanazole* . Unfortunately all attempts to induce the desired electrophilic exo cyclization on 24 even with mild protic acids have resulted in fragmentation of the sugar portion, giving back the indolocarbazole 4. Danishefsky et. al.⁶ experienced similar frustration in their handling of analogus enolic intermediate. They had to resort to nucleophlic iodo cyclization followed by reduction, to realize a compound like 7, in modest yield. Related considerations have forced us to seek alternatives and **such** refinements leading to the total synthesis of staurosporine are currently in progress and will be the subject of future publication.

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- 15. All new compounds were characterized by 1 H-NMR, 13 C-NMR, MS, and combustion analysis and/or high resolution MS. The chemical shifts of acetate methyl which are syn to the indolocarbazole appear at a higher field due to aniosotropy and they are highlighted where appropriate. Selected Spectroscopic Data: 5:¹H-NMR (400 MHz, CDCl3) δ 2.06 (s, 3H), 2.09 (s, 3H), 4.68 (d, J = 1.5 Hz, 1H), 4.93 (d, J = 1.6 Hz, 1H), 5.11 (m, 2H), 5.42 (dd, J = 2.3, 2.6 Hz, 1H), 6.57 (dd, J = 5.2, 2.0 Hz, 1H). 9:m.pt=224-227°C; ¹H-NMR (400 MHz, CDCl3) δ 1.66 (m, 2H), 2.2-2.5 (m, 4H), 6.69 (d, J = 3.8 Hz, 2H), 7.3-7.5 (m, 6H), 7.92 (s, 2H), 8.19 (d, J = 7.8 Hz, 2H). 10B: m.pt.=135-1370C; Rot.[α]D -73.40 (c=0.134, MeOH); ¹H-NMR (400 MHz, CDCl3) δ 1.74 (s, 3H), 2.05 (s, 3H), 2.98 (ddd, J = 15.08, 2.3, 1.0 Hz, 1H), 3.3 (ddd, J = 15.08, 5.95, 5.95 Hz, 1H), 3.8 (dd, J = 12.5, 2.18 Hz, 1H), 4.05 (ddd, J = 10.67, 3.17, 2.18 Hz, 1H), 4.3 (dd, J = 12.5, 3.17, 1H), 5.46 (dd, J = 10.67, 4.46 Hz, 1H), 5.58 (ddd, J = 5.95, 4.46, 2.3 Hz, 1H), 6.6 (d, J = 5.95 Hz, 1H), 7.3-8.2 (m, 10H). 10C: m.pt.=220-222°C; Rot.[a]p + 85.50 (c=0.55, MeOH); ¹H-NMR (400 MHz, CDCl₃) δ 1.58 (s, 3H), 2.3 (s, 3H), 2.9 (appt. d, J = 15.3 Hz, 1H), 3.37 (ddd, J = 15.26, 6.16, 6.16 Hz, 1H), 3.59 (dd, J = 11.88, 6.88 Hz, 1H), 3.65 (dd, J = 11.88, 6.88 Hz, 1H), 4.25 (ddd, J = 6.88, 6.88, 3.05), 5.14 (m, 1H), 5.27 (m, 1H), 6.58 (d, J = 6.16 Hz, 1H), 7.3-8.2 (m, 10H). 11B: m.pt.=256-258^oC; Rot.[α]_D-97.6^o (c=0.125,CHCl3); ¹H-NMR (400 MHz, CDCl3) δ 0.8 (d, J = 7.5 Hz, 1H), 2.28 (s, 3H), 2.87 (br. d, J = 15.3 Hz, 1H), 3.36 (ddd, J = 15.3, 6.1, 6.1 Hz, 1H), 4.18 (m, 1H), 5.13 (m, 2H), 6.58 (d, J = 6.1 Hz, 1H), 7.3-8.2 (m, 10H). 11C: m.pt.= 278-280°C; Rot.[α]_D +73.6° (c=0.14,CHCl3); ¹H-NMR (400 MHz, CDCl3) δ 1.2 (d, J = 6.1 Hz, 3H), 1.76 (s. 3H), 2.96 (ddd, J = 15.06, 5.93, 1.22 Hz, 1H), 3.26 (ddd, J = 15.06, 5.93, 5.93 Hz, 1H), 3.96 (m, 1H), 5.14 (dd, $J = 10.3$, 4.35 Hz, 1H), 5.52 (ddd, $J = 10.3$, 5.93, 1.22 Hz, 1H), 6.52 (d, $J = 5.93$, 1H), 7.2-8.2 (m, 10H). 13A: m.pt.=151-153°C; Rot.[α]_D +154.2° (c=0.184,CHCl3); ¹H-NMR (400 MHz, CDCl3) 8 1.88 (m, 1H), 1.91 (s, 3H), 2.36 (s, 3H), 2.56 (ddd, $J = 11.8$, 11.8, 10.8 Hz, 1H), 4.23 (appt. d, J = 13.1 Hz, 1H), 4.66 (appt. d, J = 13.1 Hz, 1H), 5.35 (appt. dd, J = 11.8, 4.0 Hz, 1H), 5.6 (br. S, 1H), 6.02 (dd, J = 10.8, 2.0 Hz, 1H), 7.2-8.2 (m, 10H), 9.8 (s, 1H, NH). 14A: m.pt.=202-204 $^{\circ}$ C; Rot.[α]_D-95.3° (c=0.286,CHCl3);¹H-NMR (400 MHz, CDCl3) δ 1.9 (m, 1H), 1.94 (s, 3H), 2.06 (s, 3H), 2.36 (s, 3H), 2.56 (ddd, J = 12.5, 12.5, 11.29 Hz, 1H), 4.42 (dt, J = 6.5, 1.05 Hz, 1H), 4.53 (m, $2H$), 5.35 (ddd, $I = 12.2$, 4.88, 3.05Hz, 1H), 5.64 (d, $I = 3.05$ Hz, 1H), 6.11 (dd, $J = 11.29$, 2.14 Hz, 1H), 7.2-8.2 (m, 10H), 9.78 (s, 1H, NH). 19: m.pt: 221-223°C; ¹H-NMR (400 MHz, CDCl3) 8 2.75 (d, $J = 13.8$ Hz, 1H), 3.42 (s, 3H), 3.46 (m, 1H), 4.92 (s, 1H), 5.40 (d, $J = 6.8$ Hz, 1H), 6.94 (d, $J = 7.3$ Hz, 1H), 7.3-8.2 (m, 10H). 21: 1H-NMR (300 MHz, DMSO-d6) 8 1.9 (m, 1H), 2.5 (m, 1H), 3.7 (m, 2H), 3.93 (br. t, J = 6.2 Hz, 1H), 4.04 (br. s, 1H), 4.14 (br. m, 1H), 4.83 (t, J = 5.6 Hz, 1H, D₂O exch.), 5.18 (d, J = 5.25, exch.),7.2-8.2 (m, 10H), 11.56 (s, 1H, NH, D₂O exch.). 24: m.pt.=169-171^oC; Rot.[a]_D -185.8^o (c=0.12,CHCl3); ¹H-NMR (400 MHz, CDCl₃) δ 1.94 (s, 3H), 1.97 (m, 1H), 2.32 (s, 3H), 2.76 (ddd, J = 11.36, 11.3, 10.84 Hz, 1H), 5.14 (d, J = 1.4 Hz, 1H), 5.27 (br. S, 1H), 5.37 (ddd, J = 10.84, 4.66, 3.12), 6.01 (d, J = 3.12 Hz, 1H), 6.2 (dd, J = 11.36, 2.36 Hz), 7.2-8.2 (m, 10H), 9.45 (s, 1H, NH).

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